

MEDICINAL EFFICACY OF EDIBLE MACRO FUNGUS Ganoderma applanatum (PERS.) PAT. – A REVIEW

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INTRODUCTION

Improper nutrition and modern lifestyle are the two common reasons for the growth of diseases and disorders all over the world (Kozarski *et al.*, 2015). It has been estimated by the World Health Organization (WHO) that about 80% of the earth's inhabitants rely on traditional medicine for their primary health care (Krishnaiah *et al.*, 2011). Since beginning of mankind nature is the most important source of medicines, and the bioactive compounds produced from traditionally used living organisms have been used directly as drugs or as lead compounds for drug development. Medicinal mushrooms have the potentiality to become real drugs of traditional and evidence-based medicine (Lindequist *et al.*, 2014).

The mushrooms have been defined as a potential source of dietary fiber, high quality of proteins, essential amino acids, as well as the numerous vitamins and minerals, polysaccharides, glycoproteins, unsaturated fatty acids, phenolic compounds, ergosterols, lectins etc. (Gaoxing et *al.*, 2018). In recent years innumerable studies have been focusing on the therapeutic benefits of mushrooms, including the immune boosting activity, anticancer, anti-inflammatory, antioxidant, antifungal, antibacterial, antiviral and many other medicinal properties (Rathore et *al.*, 2017; Gaoxing et *al.*, 2018).

It is estimated that nearly 1,40,000 mushroom species are found all over the world and among them only 5 percent could profit the mankind. Most edible and certain non-edible, non-consumptions mushrooms have been recognized to improve health from several thousand years ago (Bell *et al.*,2022). Traditional mycological knowledge of most Indian

ABSTRACT

Medicinal mushrooms are used in traditional medication. They are easily available and cost effective source for the people of rural and tribal areas. In the present review, an attempt has been made to congregate the taxonomic, mycochemical and pharmacological studies done on an important medicinal mushroom *Ganoderma applanatum*. Extensive experimental studies prove that *Ganoderma applanatum* possesses antibacterial, antifungal, antiviral, antioxidant, anticancer, anticholestromic, antidiabetic, nephroprotective, hepatoprotective, hypouricemic, immunostimulatory properties, which help it to play role in prevention and treatment of many disease. Therefore, it is worthwhile to review therapeutic properties of *G. applanatum* to give an overview of its status to scientist both modern and ancient.

ethnic groups has proven to be extensive and profound, consuming nearly 283 species of wild mushrooms as food and medicine out of the 2,000 species (Choudhary et *al.*,2015).

Mushrooms belonging to genus Ganoderma, commonly known as Reishi mushrooms, is the largest genus in the order Polyporales with more than 400 records, are known for their medicinal importance in the Asian continent. Boundless traditional medicinal uses of Ganoderma species have attracted the pharmaceutical industry recently (Loyd et al., 2018). Several studies reported that Ganoderma applanatum has many medicinal properties such as antibacterial, antiviral, antitumor, antifibrotic, antiobesity, antioxidative, and immunomodulatory (Luo et al., 2017 Gao et al., 2019). The goal of the present review is therefore to crystalize the knowledge of the effectiveness of Ganoderma applanatum as a possible natural oral hypoglycemic, hypolipidemic, and hepatoprotective agent, or functional food, in reducing hyperglycemia along with other vascular complications reported by different workers apart from other aspects of medicinal efficacy of the species. Ganoderma mushrooms are important medicinal mushroom, being used in clinical, pharmaceutical, and nutritional industries for the treatment of various diseases such as migraine, hypertension, asthma, hepatitis, cancer, and cardiovascular problems (Mawar et al., 2020).

This review covers bioactivities of medicinal mushroom *Ganoderma applanatum* from different countries of the world. In this context, we aimed to give an overview and summarization of the nutritional, therapeutic benefits and other bioactivities of *G. applanatum* in the present review. The

review including the bioactive nutraceuticals in *G. applanatum* and their functioning mechanisms, which could be conducive to the better understanding of the correlations between the *G. applanatum* consumption and health improvement.

Taxonomic and medicinal history

The genus *Ganoderma* was established as genus in 1881 with with single species *G. lucidum* (Curtis:Fr.) P. Karst from England as the type species by Finnish mycologist Peter Adolf Karsten and before Karsten this taxon was characterized as *Boletus lucidum* by Curtis in 1781 (Karsten, 1881; Palanna et al., 2020). The mushroom *G. applanatum* was first reported by German mycologist Christiaan Hendrik Persoon who made additions to Linnaeus mushroom taxonomy. He gave it the binomial scientific name *Boletus applanatus* and this polypore was transferred to genus *Ganoderma* in 1887 by the French mycologist Narcisse Theophilr Patouillard (First Nature, 2021).

From ancient time the mushroom of genus *Ganoderma* has been used as traditional medicinal supplement in some Asian countries such as China, Japan and India. The main product is the dried fruit body of *G. applanatum* used as traditional Chinese medicine and it has also been reported as a medicinal farming crop and is used as a flavor enhancer in Asian cuisine because of its rich mushroom flavor. In Japan it is known as *kofuki-saru-no-koshikake* and and in China as *shu-she-lingzhi* (Asian Anticancer Material Database, 2021). No ancient time period had been mention for the traditional medicinal uses of *G. applanatum*.

Lingzhi has been recognized as a medicinal mushroom for

over 2000 years, and its powerful effects have been documented in ancient scripts (Wasser *et al.*,2005). In the supplement to *Classic of Materia Medica* (502-536 AD) and the *Ben Cao Gang Mu* by Li Shin-Zhen, which considered to be the first pharmacopoeia in China (1590 AD; Ming dynasty), the mushroom was attributed with therapeutic properties, such as tonifying effects, enhancing vital energy, strengthening cardiac function, increasing memory, and antiaging effects (Wachtel-Galor *et al.*,2011).

Classification (Anand and Chowdhry, 2013; CABI, 2020)

Domain: Eukaryota

Kingdom: Fungi Phylum: Basidiomycota Subphylum: Agaricomycotina Class: Agaricomycetes Subclass: Agaricomycetidae Order: Polyporales Family: Ganodermataceae Genus: Ganoderma Species: Ganoderma applanatum Binomial name: Ganoderma applanatum (Pers.) Pat.

Distribution and Morphology

The mushroom has been reported from different countries of Asia, Europe, North America and South America. Thus their distribution can be considered cosmopolitan (CABI, 2020). In

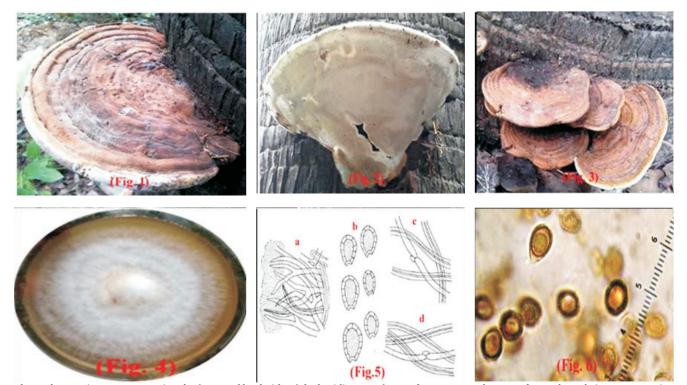


Photo plate 1: Fig. 1, 2 representing the front and back side of the basidiocarp of *G. applanatum* (Dandapat *et al.*, 2019b) and Fig. 3 representing basidiocarp of *G. applanatum* in groups; Fig. 4 diagrammatic representation of the cultured mycelia of *G. applanatum*; Fig. 5 showinghyphae of the cutis (a), basidiospores (b), tube layer hyphae (c) and context hyphae (d) of *G. applanatum* (Saha *et al.*, 2018); Fig. 6 microscopic view of spores of *G. applanatum* (Kuo M, 2018).

India this taxon was previously reported from Maharashtra, Dehradun, Punjab, Assam and Jharkhand (Saha et al., 2018; Dandapat et al., 2019). The macrofungus is saprobic, perennial but releasing spore in summer and autumn. *G. applanatum* grows alone or in groups on dead logs of hardwood, sapwood and conifers, also on living trees (Anand and Chowdhry, 2013).

The term *Ganoderma* is derived from the Greek word *ganos*: shining and *derma*: skin.The mushroom *G. applanatum* has been identified on the basis of morphology and reported from time to time. The bases of morphological identification are basediocarp, hyphal system and spores. The report on identification of *G. applanatum* by different workers is presented in Table-1 and Photoplate-1includes the list of identified structure of *G. applanatum*.

microelements like Mn, Zn, Co, Mo, Ni and Cu are also required and they are generally part of enzymes and cofactors. Microorganisms also require growth factors, which are organic compounds (Basu et *al.*,2015)

A culture media used for the *in vitro* culture of fungal mycelia is composed of four basic components- essential elements and minerals, an organic supplement supplying vitamins and amino acids, a source of fixed carbon and a gelling agent (Ajdari *et al.*,2011; Kumar and Mina, 2019). In addition, various types of C and N sources could be utilized by fungi due to their ability in secreting various enzymes for the degradation of the polymers into small molecules (Lee *et al.*, 2007). This means that the type and concentration of C and N sources, C/N ratio, and vitamins are the important factors in medium formulation for the enhancement of fungal growth

Table 1:	Description of	G. applanatum	basediocarp.	hyphae and	spores.

SI. Basediocarp	Hyphal system	Spore	References
No.			
 Dull grey, grey-brown to brown, fan shaped, hard woody in texture. Size: 5.7-6.2cm broad × 5.5-11 cm thick. Pores: 4.2-5.8 per mm, white.Multi-seried tubes: 3.7-14 mm long, separation of a thin tissue layer. 	No description	Brown, elliptical, blunt at the distal end, thick-walled, ornamented with minute spines.	Anand and Chowdhry, 2013
2 Sessile, dull, non-laccate, woody, applanate. 50–86 × 30–35 mm in diameter and 35–44 mm thick. Upper surface greyish to reddish- brown with concentric zonation, whitish pore surface.	Trimitichyphal system. Yellowish-brown generative hyphae:thin walled, 4.5–5.5 ¼m wide, clamp connection present at septa.Dark brown branched skeletal hyphae: 4.5–6 ¼m wide, and the hyaline binding hyphae: branched, 1.7–2.5 ¼m wide.	Basidiospore:double walled, truncate $7.5-9 \times 3.5-4.5 \%$ mdiameters., Exospores: hyaline, thin walled.Endospore: pale yellow to dark brown, elongated ridges	Saha et al., 2018
 Semicircular, 10–30 cm across, 8–14 cm deep, dull, unvarnished outer crust surface, furrowed in zones ,brownish to grayish brown. white pore surface, dirty brown in age, with 4–6 tiny circular pores per mm. 	The older layers of tube often stuffed with white mycelial material.	Ellipsoid, a truncated end, double- walled, brown to orangish brown, 6–9 x 4–5 µm	Kuo M., 2018
4 15 to 50cm across and 5 to 10cm thick, off-white margin and a brown top.Lower surface with small round pores, typically five per mm, are white when the fruiting body is young, turning brown with age o when bruised.		Brown, Ellipsoidal to ovoid, truncate at one end, smooth,6.5-8.5 x . 4.5-6¼m	Paul <i>et al.,</i> 2008; First Nature, 2021.
5 3-30 cm wide × 5-50 cm long × 1-10 cm thick, hard as leather, woody. The tubes are 4-12 mm deep and terminate in pores that are round with 4-6 per millimetre.	No description	Brown spores were released from the pores on the underside of the fruiting body.	Ginns J, 2017; Meuninck J. 2017
6 Semicircular, 13 cm in diameter,wrinkled zones of brownish to greyish-brown colour at outer surface and the lower surface is white.	No description	No description	Dandapat et al., 2019b

Culture and Growth

Microbial culture media can be of different type, depending on the nutritional growth requirements of the microorganisms. Microorganisms require about 10 macro elements namely C, O, H, N, S, P, K, Ca, Mg and Fe. The first six components are used in the synthesis of Carbohydrates, Lipids, Proteins and Nucleic acids and the remaining four exist in the cell as cations and play a variety of roles. In addition to macroelements, and sporulation. Although potato dextrose agar (PDA) and malt extract agar (MEA) are generally known as the most common media for growth and sporulation of fungi (Carvalho *et al.*, 2005) they are only suitable for laboratory scale and might not be economic at large-scale production. However microorganisms flourish at different environments and have some environmental growth requirements like pH, osmotic conditions, light and temperature. Woo-Sik Jo *et al.* (Woo-Sik *et al.*, 2009) conducted an experiment to obtain basic information regarding the mycelial culture conditions of *Ganoderma applanatum*. They focused mainly on the effects of pH, temperature, culture media, and effects of nutrient sources such as carbon, nitrogen, carbon/ nitrogen ratio, vitamins, organic acid and mineral salt on the growth of *G. applanatum* mycelia. They reported 6-9 pH, 25-30°C temperature, PDA, YMA (yeast malt agar), MCM (mushroom minimal media) and dark conditions showed maximum growth of *G. applanatum* mycelia. They also found among the used 10 carbon sources mannose, among the 13 nitrogen sources yeast extract, 5:1 C/N ratio, vitamins like thiamine-HCl and biotin, organic acid like succinic acid and lactic acid, salts like MgSO₄, 7H2O, KH₂PO₄ and NaCl were showed excellent growth of *G. applanatum* mycelia.

In a similar work on mycelial growth of *G. applanatum* for exo-polymer production, (Yong *et al.*,2009) it was reported that the growth of *G. applanatum* mycelia was highest and a pH of 4 to 6. Glucose served as a main carbon source among the studied five different carbon sources (glucose, maltose, arabinose, mannose and molasses) and corn steep powder served as main nitrogenous source among other studied nitrogen sources (yeast extract, peptone, meat extract, malt extract) for the maximum production of mycelia (10.65 g/L to 11.87 g/L). It was also reported maximum yield of mycelia (18.0 g/L) was achieved at the end of log phase (12 days).

Mycochemistry of G. applanatum

Woody mushrooms normally grow as parasites or saprophytes on living trees or sometimes associated with logs of wood and tree stumps. Species of Ganoderma are well known to cause different types of rots by decomposing lignin as well as cellulose and other related polysaccharides in both angiosperm and gymnosperm hosts (Singh et al., 2014). Some species of Ganoderma have been reported to contain different chemical constituents such as polysaccharides, proteins, amino acids, fatty acids, terpenoids, steroids, alkaloids, phenolic compounds, etc. (Paterson, 2006; Boh et al., 2007) with anti-inflammatory, anti-tumor, anti-oxidant, immun omodulatory, anti-diabetic, anti-viral, anti-bacterial, anti-fungal properties (Paterson, 2006; Kao et al., 2013; Singh et al., 2014). Production of secondary metabolites is a common feature of fungi. It consists of a relatively small number of enzymological processes which convert a few important intermediates of primary metabolism into a wide range of products (Bu'Lock. 1961).Furthermore, fungal secondary metabolites are derived from a few common biosynthetic pathways which branch off the primary metabolic pathways and are often produced as families of related compounds, often specific for a group of organisms (Dewick P.M., 2009). The MVA and MEP pathway, the shikimic acid pathway, the acetate pathway are the major pathways for the synthesis of fungal secondary metabolites such as terpenoids, aromatic amino acids and phenylpropanoids, polyketides respectively. Fungal alkaloids are commonly produced from amino acid through terpenes or acetate pathway (Hansoon D., 2013). Qualitative biochemical screening of different solvent extracts of G. applanatum fruiting body were carried out and found the extracts of this mushroom carry wide range of primary as well secondary metabolites such as carbohydrates, proteins, amino

acids, lipids, steroid, terpenoids, alkaloids, phenolic compounds, tannins, flavonoids, saponins, glycoside etc (Singh et al., 2014; Nagaraj et al., 2013; Dandapatet al., 2019b). Two new benzopyran-4-one derivativesapplanatin A $(C_{16}H_{16}O_6)$, B $(C_{16}H_{18}O_6)$ and a ganoderma aldehyde commonly known as ganoderenic acids were isolated and identified by column chromatography and ¹³C NMR from chloroformmethanol extract of fruiting bodies of the fungus G. applanatum (Wang et al., 2007). Palmitic acid is a major antibacterial compound that was isolated, quantified by HPLC and TLC from methanolic extract of *G. applanatum* and the molecular structure of the compound was determined by Mass and ¹H NMR spectroscopy (Moradali, 2008).By using GC-Mass and $H_1 NMR$ bioactive chemical compounds G1 ($C_{20}H_{34}O_4$) with chemical structure [(19,19a-dihydroxy-2-methyl-2,3,4, 5,6,7,8,9,10,11,12,13,14,15 tetrad ecahydrobenzo[b] oxacycloheptadecin-17(19aH] belongs to tannin group and another compound G2 ($C_{21}H_{28}O_2$) with molecular structure[(2-(2-(2,5- dihydroxyphenyl) ethylidene)- 11-hydroxy-6,10dimethylundeca-5,9-dienoic acid] belongs to terpenoides group were isolated and identified from the mycelia culture of G. applanatum which showed potent bactericidal activity (Muhsin et al., 2011).

A chemical analysis of ethanolic extract of mushroom *G*. *applanatum* was carried out through gas chromotaghrophy coupled to mass spectrometry and first time reported six new compounds:4-metil-ergosta-22-en-3 α -ol; 4,4-dimetil-ergosta-5,7,25(26)-trien-3 -ol; ergosta-5,7,9,24(28)-tetraen-3 α -ol; 4,4-dimetil-ergosta-7,24(28)-dien-3 -ol; 17-isopropil-"^{5,7}-3odroxiandrostadieno; and estigmasta-5,22-dien-3 a-ol (Guzman et al., 2013). In another study GCMS analysis of methanolic extract of *G. applanatum* was performed and identified some principal components a-terpinene, Dlimonene, cis-2-methyl-4-pentylthiane-s, s-dioxide, α -cymene and -terpinolene which possessed cytotoxic potential on different tumour cells (Hakkim et al., 2016).

Quantitative estimation of secondary metabolites of mushroom G. applanatum has been done from different regions of world and the results showed variation in the amount and composition of secondary metabolites.Nagaraj et al.(2014), estimated phenol (71 \pm 0.02 μ g/mg) and flavonoid $(45\pm0.01\mu g/mg)$ in high quantity compared to tannin $(12.82\pm0.02\mu g/mg)$, saponin $(5.91\pm0.01\mu g/mg)$ and steroid $(13.45 \pm 0.01 \mu g/mg)$ from methanolic extract of *G*.applanatum. In a similar study Rajoriya et al. (2015) quantified proximate secondary metabolite from *G. applanatum* fruiting body. They found phenol (11.60 \pm 0.20 mg/g), caretonoid(7.43 \pm 0.29 mg/ g) and alkaloid (4.40 $\pm\,0.25$ mg/g) in high amount compared to the other trace metabolites such as -carotene, lycopene, ergosterol, tannin, ascorbic acid and flavonoids. Mohammadifar et al. (2020) also performed a comparison study between antioxidant activity and bioactive compounds of G. applanatum and G. lucidum and reported the extract of G. applanatum contain 6.70 mg/g phenolic compound and 1.37 mg/g flavonoid compound. They also quantified terpenoid acids, ursolic acid and oleanolic acid in trace and 1.55mg/g betulinic acid by HPLC method.

Pharmacological and medicinal activities

Antimicrobial activity

G. applanatum possess a wide range of bioactive compounds and they showed broad spectrum antibacterial, antifungal and antiviral activities. The mushrooms showed antibacterial activities against both gram-positive and gram-negative bacteria (Gao *et al.*,2005). Artur *et al.*,(1999), studied the bactericidal activity of isolated sterols 5 α -ergost-7en-3 α -ol, 5 α -ergost-7,22-dien-3 -ol, and 5,8-epidioxy-5 α , 8 α -ergost-6,22-dien-3 -ol against pathogenic ATCC and MIP strain bacteria Bacillus cereus, Corynebacterium diphtheria, E. coli, *P.* a*eruginosa*, *Staphylococcus aureus*, *Staphylococcus saprophyticus*, and *Streptococcus pyogenes*. They reported gram-positives bacteria were more sensitive (MICs of 0.003 to 2.0 mg/mL; MBCs of 0.06 to 4.0 mg/mL; MBCs of 2.0 to > 4.0 mg/ mL).

The crude extracts of G. applanatum are also well known for their antimicrobial activities. A broad range of antibacterial and antifungal activity of petroleum ether, chloroform, methanolic and aqueous extracts of G. applanatum fruiting body were screened against MTCC pathogenic bacteria S. aureus, B. subtilis, E. coli, P. aeruginosa, K. pneumonia, S. typhi and four dermatophytic fungi C. albicans, C. indicum, T. rubrum and M. gypseum by agar well diffusion method (Nagaraj et al., 2013). Among the studied bacteria S. typhi showed lowest inhibitory effect against all the extracts (only 8 \pm 0.24 mm zone of inhibition against methanolic ectract) and S. aureus showed highest sensitivity $(15\pm0.21 \text{ mm to } 8\pm$ 0.21 mm zone of inhibition) against all the studied extracts. Among the studied fungal strains C.albicans (10 ± 0.23 mm to $5\pm$ 0.2mm ZOI) and C.indicum (7 \pm 0.18 mm to $5\pm$ 0.1 mm ZOI) showed high inhibition to all the extracts.

Monika et al. (2014), studied the antibacterial activity of isolated exopolysaccharide from stationary cultures of *G. applanatum* against *E. coli* and *S. aureus* strains. The exopolysaccharide showed 17.9mm inhibition zone of and 1mg/mL MIC against *S. aureus* but no inhibition was observed against *E. coli*.

Bactericidal activities of purified polysaccharides and hot alkali extract of wild G.applanatum were examined against some other bacterial species such as Listeria monocytogenes, Geobacillus stearothermophilus, Enterococcus faecalis, Proteus hauseri, Salmonella enteritidis, Shigella sonnei and Yersinia enterocolitica(Klaus et al., 2016) and found the alkali extract (MIC: 0.039 mg/mL to 1.25 mg/mL; MBC: 1.25 mg/mL to 10.0 mg/mL) was most effective than the partially purified polysaccharide (MIC: 0.156 mg/mL to 5.0 mg/mL; MBC: 2.5 mg/mL to 10.0mg/mL). Bacterial growth inhibitory activity of methanol, ethanol, chloroform and aqueous extracts G.applanatum has been studied by agar well diffusion and broth dilution method against urethitis, cystitis, prostatitis and pneumonia causing bacteria Proteus mirabilis (Dandapat et al., 2016) and has been found that all the extracts, in both the methods adopted, showed 100% inhibition indicating $1.98 \pm$ 0.01 to 3.02 \pm 0.03 mm ZOI with 800 μ g/mL MIC.

It is reported that antimicrobial compounds such as terpenes, lectins, polysaccharides etc. act on the bacterial cytoplasmic membrane to make them more permeable for autolysis (Lin and Chou, 1984; Yang et al., 2002; Qureshi et al., 2010). It has been experimentally reported the *Ganoderma* mycelia and fruiting body extracts inducing protein leakage within bacterial cells due to their rich content of flavonoids and nucleobases (Mishra et al., 2018). During pathogenic invasion extract of medicinal mushrooms induced generation of free radicals such as reactive oxygen species, peroxide radicals, and superoxide radicals etc. within bacterial cells, leading to cellular membrane damage as well damage intra cellular components such as DNA, intermediate enzymes involve in membrane synthesis etc (Zhang et al., 2015; Mishra et al., 2018).

Viral infections are amongst the most common diseases affecting people worldwide. New viruses emerge all the time and presently we have limited number of vaccines and only few antivirals to combat viral diseases. Fungi represents a vast source of bioactive molecules, which could potentially be used as antivirals in the future (Rikka *et al.*, 2018). Antiviral activity of *G. applanatum* has not been widely explored. Elkhateeb *et al.*, (2020) studied the antiviral activity of methanolic extract *G. applanatum on* simian rotavirus SA-11 strains by MTT assay method and they found the extract exhibited Tl (CC_{50}/IC_{50}) = 3.4 against virus infection.

Antioxidant activity

Antioxidants are any substance, that delays, prevents, or removes oxidative damage to a target molecule (Halliwell B., 2007). Antioxidant compounds are broadly classified into natural or primary and synthetic or secondary antioxidants. They are the chain breaking molecules which react with free radicals and convert them into more stable products. They are mainly derived from the natural sources such as antioxidant minerals, vitamins, secondary metabolites of plants and higher fungi etc. Secondary antioxidants are compounds that perform the function of capturing free radicals and stopping the chain reactions and include compound such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), tertiary butyl hydroquinone (TBHQ), nordi hydroguaiaretic acid (NDGA) etc (Carocho et al., 2013; Rasheed et al., 2019).

Oxidative stress is a phenomenon caused by an imbalance between production and accumulation of free radical species (reactive oxygen species, reactive nitrogen species, reactive sulfur species, reactive carbonyl species *etc.*) in cells and tissues and their intensity wise ranging from physiological oxidative eustress to excessive and toxic oxidative distress (Yan L.J., 2017). Free radicals are generated from both endogenous and exogenous sources. Immune cell activation, inflammation, ischemia, infection, cancer, excessive exercise, mental stress, and aging are all responsible for endogenous free radical production. Exogenous free radical production can occur as a result from exposure to environmental pollutants, heavy metals, certain drugs, chemical solvents, cigarette smoke, alcohol, and radiations *etc.* (Valkoet *al.*, 2006; Valko *et al.*, 2007).

At low or moderate concentrations, free radicals play several beneficial roles such as elimination of intracellular pathogens, tumor cells, and play regulatory role in intracellular signaling cascades (Pizzino *et al.*, 2017). Oxidative distress is associated

with several patho-physiological conditions and one on the leading cause of cardiovascular diseases, chronic obstructive pulmonary disease, chronic kidney disease, neuro degenerative diseases, and cancer including sarcopenia and frailty etc. (Liguori et al.,2018).Chemical composition and antioxidant potential of mushrooms have been intensively studied and found edible mushrooms might be used directly in enhancement of antioxidant defenses through dietary supplementation to reduce the level of oxidative stress. Wild or cultivatedmushrroms have been related to significant antioxidant properties due to their bioactive compounds, such as polyphenols, polysaccharides, vitamins, carotenoids and minerals (Kozarski et al.,2015).

In vitro antioxidant activity of G. applanatum petroleum ether, chloroform, methanol and aqueous extract were determined by DPPH scavenging assay, reducing power assay, metal chelating assay, hydroxyl radical scavenging assay, superoxide radical scavenging assay. The methanolic extract showed higher DPPH radical scavenging activity (IC₅₀:135.13 μ g/mL), ferric reducing activity (0.85%), Fe²⁺ chelating activity (IC₅₀: 50.84 μ g/mL), compared to the other extract, hydroxyl radical scavenging activity (IC_{50}: 104.89 $\mu g/mL$ to 112.78 $\mu g/mL),$ superoxide radical scavenging activity (IC₅₀: 111.94 μ g/mL and 120 μ g/mL) compared to the other extract (Nagaraj et al., 2014). Rajoriya et al. (2015), studied and reported G. applanatum methanolic extract showed 91.64% DPPH radical scavenging activity with $IC_{50} = 6mg/mL$. Antioxidant activity of exoploysaccharide of G. applanatum mycelia was studied and reported, the exopolysaccharides of G. applanatum strain effectively scavenged hydroxyl radicals (63.9±0.1%) and superoxide anion radicals (58.8 ± 0.1) (Liu et al., 2015).

Anticancer activity

Cancer is a major public health problem, being one of thehighest causes of death in both men and women in developed as well as developing countries (Baliga et al., 2012; Rahman et al., 2014). Approximately 12.7 million new cancer cases and 7.6 millioncancer deaths occurred near 2008. By the year 2020, predictions report that the incidence of cancer may increase 3-fold, with a disproportionate rise in cancer cases and deaths in developing countries (Rahman et al., 2014).Cancer occurs by successive mutations in genes that changes cell functions. Chemical compounds have an obvious role of forming gene mutations and cancer cells but other factors such as smoking, viruses, bacteria, and radiation rays are other carcinogenic factors, comprising about 7% of all cancers (Parkin D.M., 2006; Rahman et al., 2014). In general, cancer disrupts cellular relations and results in the dysfunction of vital genes and this disturbance is affective in the cell cycle, and leads to abnormal cell proliferation (Seto et al., 2010; Rahman et al., 2014).

Cancer in the broader sense refers to more than 277 different types of cancer disease. In men commonly the highest percentage of cancer occurs in the prostate, lung and bronchus, colon and rectum, and urinary bladder, and in women most prevalence cancer occur in the breast, lung and bronchus, colon and rectum, uterine corpus and thyroid (Siegel *et al.*, 2016;Rahman *et al.*, 2014) . For children, the highest percentage types of cancer are blood cancer, and cancers

related to the brain and lymph nodes (Rahman et al., 2014). Chemical drugs for cancer treatment, such as cisplatin and cyclophosphamide, can cause side effects, such as nephrotoxicity, which are detrimental to the quality of life of patients (Aguirre et al., 2013). In addition to this toxicity, the resistance of some cancer cells to treatment has led to the need for the evaluation of alternative approaches (Yu et al., 2018).

It has been reported the anti-cancer compounds of mushroom play crucial role as reactive oxygen species inducer, mitotic kinase inhibitor, anti-mitotic, angiogenesis inhibitor, topoisomerase inhibitor, leading to apoptosis, and eventually checking cancer proliferation (Patel *et al.*, 2012). The main *Ganoderma* species are *G. lucidum*, *G. sinensis*, *G. applanatum*, *G. tsugae*, *G. atrum*, *G. formosanum*, *G. microsporum*, and *G. atrum*. They have been studied and broadly explored for anticancerous and other medicinal properties (Yu *et al.*, 2018).

Antitumor effect of exo-polymer produced by G. applanatum was investigated using sarcoma-180 bearing mice and significant growth inhibition of solid tumor and increase natural killer (NK) cell activity as well phosphatase activity was found (Yong-Tae et al., 2008). Antitumor activity of extracellular polysaccharide from cultured mycelia of G. applanatum was investigated in vitro by MTT assay against cervical carcinoma cell lines SiHa and CaSki (Monika et al., 2014). A significant cytotoxic activity with 42.8% and 34% decrease in cell viability of SiHa carcinoma at 22.88 5µg/mL and 228.5 5µg/mL concentrations of the exopolysaccharides was observed respectively. It has been reported that polysaccharides from Ganoderma species inhibited tumor growth via induction of apoptosis through mitochondrial pathways and immuno enhancement effects (Li et al., 2011). A dose dependent antiproliferative activity of terpenoid compounds from methanolic extract of *G. applanatum* was studied against triple negative breast cancer cells (MDA-MB-231) and cervical cancer cells (HEp-2) and found after 24 hours of treatment the carcinoma cells lost their adherence and morphology and were found to be necrotic (Faruck et al., 2016). Recently both in vivo and in vitro anticancer study of G. applanatum secondary metabolites was carried out and reported that the G. applanatum inhibit the proliferation and viability of colon cancer cell (Ccco-2) and decrease the volume of solid developed Ehrlich tumor by activating apoptosis through p53-independent and p53dependent path ways (Elkhateeb et al., 2018).

Anti-inflammatory activity

Inflammation is considered to be part of the complex biological response to remove injury or harmful stimuli such as pathogens, damaged cells, or irritation. Inflammation leads to many physiological symptoms such as fever, pain, and swelling, vasodilation, increased vascular permeability etc. In recent era nonsteroidal anti-inflammatory drugs (NSAIDs) are commonly used to treat inflammation but long-term use of NSAIDs induce side effects such as mucosal lesions, bleeding, peptic ulcers, intestinal perforation etc. (Sinha et al., 2013). Side effects of NSAIDs are not limited only to GI-tract but extend to serious situations such as acute renal failure, nephrotic syndrome, hypertension, and cardiovascular toxicity etc. (Meek et al., 2010).

Anti-inflammatory effects with anti-inflammatory compounds of mushrooms involve a highly diversified group of biochemicals. They include polysaccharides, terpenoids, phenolic compounds, and many other low molecular weight molecules (Elsayed et al., 2014). Ganoderma extract treatment leds to significant increases in the percentages of CD⁴⁺, CD²⁵⁺, Foxp³⁺, Treg and IL-10⁺ Breg cells, together with a reduction in the plasma concentrations of several inflammatory cytokines and down-regulated expression of the corresponding cytokine related genes in systemic lupus erythematosus include inflammation of the joints, joint pain, and edema (Cai et al., 2016).In an in vitro experiment ethanol extract of G. applanatum showed dose dependent anti-inflammatory activity. The ethanol extract of G. applanatum at a dose of 600 mg/kg showed maximum suppression of oedema (59.2%) in the rat paw. Along with anti-inflammatory activity the extract also prolonged the reaction time in mice to noxious thermal stimuli (Ede et al., 2012).

Antidiabetic activity

Diabetes mellitus is a group of physiological dysfunctions characterized by hyperglycemia resulting directly from insulin resistance, inadequate insulin secretion, or excessive glucagon secretion. Type-1 diabetes (T1D) is an autoimmune disorder leading to the destruction of pancreatic beta-cells and type-2 diabetes (T2D) is most common and is primarily a problem of progressively impaired glucose regulation due to a combination of dysfunctional pancreatic beta cells and insulin resistance (Blair M., 2016). Due to imbalance of specific receptors such as Glucagon-like peptide-1(GLP-1) receptor, peroxisomes proliferator activated (a) receptor (PPARa), beta3 (α 3) ardent-receptor some enzymes like α glycosidase, dipeptidyl peptidase IV enzyme etc. can cause diabetes mellitus (Nishita et *al.*, 2016).

In vivo antidiabetic activity of *G. applanatum* extract and secondary metabolites has been studied on streptozotocin and alloxan induced diabetic animal. Streptozotocin is a glucosamine nitrosourea compound that induces diabetes either by inactivation of the GLUT2 glucose transporter receptor of α -cells or by the destruction of α - cells (Graham et *al.*, 2011). Alloxan is a pyrimidine, structurally similar to glucose and uric acid directly disrupt α -cell membrane permeability and inhibition of tricarboxylic acid cycle and Ca²⁺- dependent dehydrogenases in the mitochondria causes ATP deficiency, cessation of insulin production, and eventual α -cell death (Schmeltz et *al.*, 2006). Hossain et *al.*, (2021) reported very effective impact on blood glucose levels in

experimental rats as presented in Table 2.

Ganoderma applanatum exo-polymer (GAE) produced by submerged mycelial cultures substantially reduced the plasma glucose levels by as much as 22.0% in streptozotocin induced diabetic rats, when compared with the control group (Yang et al., 2007). Oral glucose tolerance test is a standard procedure to identify persons at high risk for type-II diabetes (Stern et al., 2002). It is a widely utilized method which has been used to evaluate the α -cell function and insulin resistance (Stumvoll et al., 2000). C. applanatum methanolic and chloroform extracts(500mg/kg b.wt.) also significantly maintained the glucose homeostasis in alloxan induced diabetic rats by inhibiting the rise in plasma glucose levelscompared to post versus pre-prandial plasma glucose levels which is nearly similar to normal control and positive control rats. The presence of various secondary metabolites helps to restore the capacity of pancreatic α -cells, stimulating insulin secretion and glucose uptake by the peripheral tissues (Ruby et al., 2018; Hossain et al., 2021).

Hepatoprotective and Nephroprotective activity

The liver is one of the most important organs of the body and performs elementary role in the regulation of different physiological processes, and its activity is related to different vital functions, such as metabolism, secretion, and storage.Hepatic diseases continue to be among the principal threats to public health, and they are a problem worldwide

(Adewusi and Afolayan, 2010; Madrigal-Santillan et al., 2014). Hepatic disease is a term that indicates damage to the cells, tissues, structure, or liver function, which can be induced by biological factors (bacteria, virus, and parasites) and autoimmune diseases (immune hepatitis, primary biliary cirrhosis), as well as by the action of different chemicals, such as some drugs (high doses of paracetamol and antitubercular drugs), toxic compounds (CCl4, thioacetamide, dim ethylnitrosamine, D-galactosamine/lipopolysaccharide), and unguestionably, excessive consumption of alcohol (Deshwal et al., 2011; Madrigal-Santillan et al., 2014). Injury to the liver, whether acute or chronic, eventually results in an increase in serum concentrations of total bilirubin (TB), aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) (Gowda et al., 2009). In normal rats G. applanatum aqueous extract showed no significant effect on TB, AST, ALT and ALP but significant elevation in serum albumin and protein level was observed (Dandapat et al., 2019). In an experiment significantly decrease AST, ALT, ALP, total bilirubin and increase in albumin level in alloxan induced diabetic rats group was observed compared to the

Table 2: Effect of Methanol extract (MEGA) and Aqueous extract (AEGA)on blood glucose level in different groups of experimental rats (Hossain *et al.*, 2021)

Groups			Blood glucose level (mg/dl)		
Day 1	Day 4	Day 7	Day 9		
96.50 ± 3.73	99.83 ± 4.96	97.00 ± 3.16	100.17 ± 3.49		
285.83 ± 11.48^{a}	290.50 ± 9.59^{a}	303.50 ± 6.06^{a}	305.83 ± 6.05^{a}		
135.83 ± 6.79^{a}	127.83 ± 5.49^{a}	119.17 ± 4.26^{a}	108.33 ± 4.32^{a}		
129.17 ± 2.99^{a}	117.50 ± 10.03^{a}	108.00 ± 3.03^{a}	102.00 ± 3.69^{a}		
149.33 ± 4.18^{a}	133.00 ± 5.69^{a}	127.67 ± 5.96^{a}	124.33 ± 4.89^{a}		
142.17 ± 4.17^{a}	127.67 ± 4.76^{a}	124.33 ± 4.41^{a}	110.33 ± 5.82^{a}		
123.50 ± 3.73^{a}	98.83 ± 7.63^{a}	91.33 ± 4.27^{a}	86.50 ± 6.22^{a}		
	$\begin{array}{r} 96.50 \pm 3.73 \\ 285.83 \pm 11.48^{a} \\ 135.83 \pm 6.79^{a} \\ 129.17 \pm 2.99^{a} \\ 149.33 \pm 4.18^{a} \\ 142.17 \pm 4.17^{a} \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $		

Groups III, IV, V, VI, and VII compared with diabetic control.^a = p < .001, ^b = p < .01, ^c = p < .05.

diabetic control group (TB- 1.38 ± 0.18 mg/dL,ALB- 2.87 ± 0.19 mg/dL, AST- 52.33 ± 4.72 mg/dL, ALT- 126.33 ± 5.79 mg/dL, ALP- 248.83 \pm 5.46 mg/dL) when treated with 500mg/ kg G. applanatum methanolic extract (TB-1.35 ± 0.11 mg/dL, ALB-3.20 ± 0.28 mg/dL, AST-25.00 ± 3.74 mg/dL, ALT-98.00 \pm 4.38 mg/dL, ALP-224.17 \pm 3.87 mg/dL) and aqueous extract (TB-1.33 \pm 0.13 mg/dL,ALB-3.20 \pm 0.30 mg/dL, AST-30.00 ± 5.10 mg/dL, ALT-98.83 ± 3.25 mg/dL, ALP-228.00 ± 5.22 mg/dL) (Hossain et al., 2021). It has also been reported sulfated Ganoderma applanatum residue polysaccharides act as potent antifibrotic agent and reduce histopathological damages, down-regulated cytochrome P450 2E1 expression, reduced serum AST and ALT, improved the anti-oxidative and anti-inflammatory properties, inhibited TLR4/NF-kB signaling pathway, and reduced the release of inflammatory cytokines (Xinling et al., 2021).

Chronic kidney disease recognized as a major risk factor for the development of new acute kidney injury, and acute kidney injury now accepted to lead to *de novo* or accelerated chronic and end stage kidney diseases (Ferenbachand Bonventre, 2016).Chronic kidney disease is most commonly attributed to diabetes and hypertension (Coca *et al.*, 2012). Acute kidney injury (AKI) describes a sudden loss of kidney function that is determined on the basis of increased serum urea, creatinine, uric acid levels and oliguria (Gowda *et al.*, 2010;Kellum *et al.*, 2021).Dandapat *et al.*, (2019), reported the aqueous extract of *G. applanatum* showed a nephro protective activity in

nomal rat and significantly decrease serum urea $(57.62\pm0.74$ mg/dL),creatinine $(0.86\pm0.008$ mg/dL) and uric acid $(14.24\pm0.73$ mg/dL) level compared to control treatment groups of rat (SU: 62.32 ± 1.17 mg/dL; CRE: 0.93 ± 0.01 mg/dL; UA: 22.5 ± 0.70 mg/dL).

Antihyperlipidemic activity

Hyperlipidemia is a medical condition characterized by an increase in one or more of the plasma lipids, including triglycerides, cholesterol, cholesterol esters, phospholipids and or plasma lipoproteins including very low-density lipoprotein and low-density lipoprotein along with reduced high-density lipoprotein levels (Shattat G.F., 2014). Elevated levels of cholesterol (hypercholesterolemia) and abnormal lipid profiles (dyslipidemia) are important risk factors for cardiovascular disease (CVD) and the most common forms of the CVD are hypertension, coronary heart disease (CHD), stroke, and heart failure etc (Mensah and Brown, 2007; Schaiff et al., 2008).In vitro hypocholesterolemic activity of G. applanatum extract was studied and reported that the cholesterol reduction activity increases proportionally with the increase in concentration of the extract, and elongation of incubation time $(90.9 \pm 1.0\%)$ to 100.0 ± 0 % reduction after 96 hours) (Elkhateeb et al., 2020). Hossain et al., (2021), reported methanolic extract of G. applanatum significantly decrease total cholesterol, LDL cholesterol and triglyceride in extract treated diabetic rats (TC: 48.83 ± 2.79 mg/dL, LDL-C: 11.83 ± 2.32 mg/dL, Triglyceride: 81.50 ± 2.74) compared to control rats (TC: 66.83 ± 2.71 mg/ dL, LDL-C: 26.67 ± 3.01mg/dL, Triglyceride: 107.50 ± 4.14mg/dL). However a significant increase in HDL cholesterol $(31.50 \pm 1.87 \text{ mg/dL})$ of extract treated diabetic rats was observed compared to control group (28.00 \pm 1.41 mg/dL). The suppression of LDL oxidation by *G. applanatum* may reduce the serum LDL significantly. Low triglyceride levels can contribute to decreasing the availability of fatty acids for esterification (Packard et al., 2020), increased catabolism of LDL, activation of tissue lipases, and decreased production of precursors of triglycerides (McCarty, 2001; Packard et al., 2020; Hossain et al., 2021). This observed restoration of the alloxan-evoked changes in the serum lipid profile shows the protective nature and hypolipidemic effect of *G. applanatum*. The suggestion is that reductions in intestinal cholesterol and bile acid absorption after the feeding of *G. applanatum* methanolic extract and *G. applanatum* aqueous extract may represent main mechanism for hypolipidemic mechanisms of *G. applanatum*(Nagaoka et al., 2005; Hossain et al., 2021).

Hypouricemic activity

In recent years, a large body of evidence has accumulated that suggests that hyperuricemia may play a role in the development and pathogenesis of a number of metabolic, hemodynamic, and systemic pathologic diseases, including metabolic syndrome, hypertension, stroke, and atherosclerosis (Billiet et al., 2014). Hyperuricemia has long been established as the major etiologic factor in gout. It is the most well understood and described type of arthritis. Gout is characterized with reoccurring inflammatory arthritis, which is triggered by the crystallization of serum uric acid (SUA) in the joints and is often caused by hyperuricemia (Choi et al., 2005) G. applanatum ethanolic and aqueous remarkably lowered the level of serum uric acid via elevated urin uric acid levels in potassium oxonate and hypoxanthine induced hyperuricemia mice, there by confirming the uricosuria effect (Yong et al., 2018). Ethanolic and aqueous extract did not decrease the XOD activity; hence the hypouricemia effects of G. applanatum may be raised by other methods, such as uricosuria or anti npurine absorption pathways. It has been reported renal transporters, including OAT-1, GLUT-9, and URAT-1, are directly associated with the homeostasis of serum uric acid (Anzai et al., 2010; Yong et al., 2018). Both ethanolic and aqueous extract of G. applanatum increased the mRNA and protein levels of OAT-1, inducing the remarkably enhanced uric acid excretion by advancing the uric acid transportation through OAT-1. The extracts also enhanced the excretion of uric acid by inhibiting the uric acid reabsorption through GLUT-9. Further more, ethanol extract down regulated the level of URAT-1 protein (Yong et al., 2018). Notably, the upregulation of OAT-1 and down regulation of GLUT-9 and URAT-1 correlated to urine uric acid provided the evidence that the synergistic activity of OAT-1, GLUT-9, and URAT-1 by G. applanatum contributed to the uricosuria actions.

Anti- Alzheimer's activities

Alzheimer's disease (AD) has progressive neuro degenerative pathology with severe economic and social impact. AD is the dementia associated with aging, which initially targets memory and progressively destroys the functions of the brain, as the neocortex suffers neuronal, synaptic, and dendritic losses. There is currently no cure, although cholinesterase inhibitors provide effective temporary relief of symptoms in some patients (Saify *et al.*, 2014). Treatment of AD has been dominated by the use of acetylcholinesterase (AChE) inhibitors. These drugs compensate for the death of cholinergic neurons and offer symptomatic relief by inhibiting acetylcholine (ACh) turnover and restoring synaptic levels of this neuro transmitter (Rees and Brimijoin, 2003). Nowadays, drug research and development are based on the cholinergic hypothesis that supports the cognition improvement by regulation of the synthesis and release of acetylcholine in the brain (Saify et *al.*, 2014). Both methanolic and aqueous extract of *G. applanatum* showed cholineesterase inhibition activity but the methanolic extract had high acetyl cholinesterase (1.45 \pm 0.01 mg gallic acid equivalents per g extract) and butyryl cholinesterase(2.94 \pm 0.19 mg gallic acid equivalents per g extract) (Zengin et *al.*, 2015).

Immunemodulatory activity

The human body has a remarkably sophisticated immune system consisting of white blood cells and specialized immune molecules that protect the body against invading pathogens (Yuandani et al., 2021). Several cytokines also play essential roles in immune response, which consist of pro-inflammatory cytokines such as tumor necrosis factor-alpha (TNF-á), interleukin 1 (IL-1), IL-6, IL-11, IL-8, and anti-inflammatory cytokines or cytokines inhibitor such as IL-4, IL-10, and IL-13 (Yuandani et al., 2021). Cytokines as intercellular messenger molecules have several functions, and these include stimulating phagocyte migration and coordinating early responses of monocytes, macrophages, dendritic cells, and lymphocytes during inflammatory states (Shaikh, 2011).Immunomodulators are natural or synthetic substances that modulate or modify the immune response. They help regulate or normalize the immune system by either stimulating or suppressing the immune system (Kumar et al., 2012).

Immunomodulatory activity of *G. applanatum* exopolysaccharide was investigated using THP-1, an acute monocytic leukemia cell line as macrophase model (Monika et *al.*, 2014). They reported, after 24 hours of incubation the extracellular polysaccharides from *G. applanatum* stimulated secretion of IL-6 by macrophages at a level of 328.5pg/mL. The 6-hour incubation of extracellular polysaccharides at a concentration of 228.5 5µg/mL resulted in production of TNF-

by 752.17 pg/mL, representing an approximately 54-fold increase compared to the negative but a decrease in the TNF-

level to 190.52 pg/mL was observed after 24-hour incubation which was higher in comparison with the negative control.

REFERENCES

Adewusi, E.A. and Afolayan, A.J. 2010. A review of natural products with hepatoprotective activity. J. Med. Plants. Res. 4:1318-1334.

Bell, V., Silva, C.R.P.G., Guina, J., Fernandes, T.H. 2022. Mushrooms as future generation healthy foods. *Front. Nutr.* 9: 1050099

Aguirre, A., Villeda-Hernandez, J., Campos-Pena, V., Herrera-Ruiz, M., Montiel, E., Tello, I. et al. 2013. Anticonvulsant and neuroprotective effects of oligosaccharides from lingzhi or reishi medicinal mushroom, *Ganodermalucidum* (Higher Basidiomycetes). *Int. J. Med. Mushrooms.* **15:** 555-568.

Ajdari, Z., Ebrahimpour, A., Manan, M. A., Hamid, M., Mohamad, R., Ariff, A. B. 2011. Nutritional requirements for the improvement of growth and sporulation of several strains of *Monascuspurpureus* on solid state cultivation. *J. Biomedicine and Biotechnology*. 487329:

1-9.

Anand, N. and Chowdhry, P.N. 2013. First report on five hitherto unreported macro fungi from Rajouri district of Jammu & Kashmir (J&K), India. *Annals of Biological Research.* **4(5):** 62-70.

Anzai, N., Jutabha, P. and Endou, H. 2010. Renal solute transporters and their relevance to serum urate disorder. Curr. Hypertens. Rev. 6: 148-154.

Asian Anticancer Material Database. 2021. Ganoderma applanatum. http://www.asiancancerherb.info/Shu%20She.htm

Baliga, M.S., Thilakchand, K.R., Rai, M.P., Rao, S. and Venkatesh, P. 2012. *Aegle marmelos* (L.) Correa (Bael) and its phytochemicals in the treatment and prevention of cancer. Integr. *Cancer Ther.* 12(3): 187-196.

Basu, S., Bose, C., Ojha, N., Das, N., Das, J., Pal, M. and Khurana,
S. 2015. Evolution of bacterial and fungal growth media.
Bioinformation. 11(4): 182 – 184.

Billiet, L., Doaty, S., Katz, J.D. and Velasquez. M.T. 2014. Review of hyperuricemia as new marker for metabolic syndrome. *ISRN Rheumatol.* 852954.

Blair M. 2016. Diabetes mellitus review. Urologic Nursing. 36(1): 27 – 36.

Boh, B., Berovic, M., Zhang, J. and Bin, L.Z. 2007. *Ganodermalucidum* and its pharmaceutically active compounds. Biotechnol. *Annu. Rev.* **13:** 266-231.

Bu'Lock, **J.D. 1961**. Intermediary metabolism and antibiotic synthesis. *Adv. Appl. Microbiol.* **3:** 293 – 342.

CABI. 2020. Invasive species compendium. Datasheet, Ganoderma applanatum (shelf fungus). https://www.cabi.org/isc/datasheet/24923

Cai, Z., Wong, C.K., dong, J., Jiao, D., Chu, M., Leung, P.C., Lau, C.B.S., Lau, C.P., Tam, L.S. and Lam, C.W.K. 2016. Anti-inflammatory activities of *Ganodermalucidum* (LIngzhi) and San-miao-San supplements in MRL/lpr mice for the treatment of systemic lupus erythematosus. *Chin. Med.* **11(23):** 1 – 13.

Carocho, M. and Ferreira, C.F.R. 2013. A review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology.* **51:** 15 – 25.

Carvalho, J. C. et al. **2005.** Biopigments from *Monascus*: strains selection, citrinin production and colour stability. *Brazilian Archives of Biology and Technology*. **48**: 885 – 894.

Choi, H.K., Mount, D.B. and Reginato, A.M. 2005. Pathogenesis of gout. Ann. Intern. Med. 143: 499-516.

Choudhary, M., Devi, R., Datta, A., Kumar, A. and Jat. H.S. 2015. Diversity of wild edible mushrooms in Indian subcontinent and its neighbouring countries. *Recent Advances in Biology and Medicine*. 1: 69 – 76.

Coca, **S.G.,Singanamala**, **S. and Parikh**, **C.R. 2012.** Chronic kidney disease after acute kidney injury: a systematic review and metaanalysis. *Kidney Int.* **81(5):** 442 – 448.

Dandapat, S., Kumar, M., Ranjan, R. and Sinha, M.P. 2019a. Study of Impacts of *Ganoderma applanatum* (Pres.) Pat. Extract on hepatic and renal biochemical parameters of rats. *Trad. Med. J.* 24(2): 119–132.

Dandapat, S., Kumar, M., Ranjan, R. and Sinha, M.P. 2019b. Acute and sub-acute toxicity of *Ganoderma applanatum* (Pres.) Pat. Extract mediated silver nanoparticles on rat. *Notulae Scientia Biologicae*. **11(3)**: 351 – 363.

Dandapat, S., Kumar, M., Ranjan, R., Sinha, M.P. 2019. Study of impacts of *Ganodermaapplanatum* (Pres.) Pat. Extract of hepatic and renal biochemical parameters of rats. *MajalahObt. Tradisional*. 24(2): 119 – 132.

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Dandapat, S., Sinha, M.P., Sarma, T.C. 2016. Antibacterial activity of wild medicinal mushroom: *Ganodermaapplanatum* (Pers.) Pat. *The Ecoscan.* 9(Spl. Issue): 837 – 843.

David Hansson. 2013. Structure and biosynthesis of fungal secondary metabolites: Studies of the Root Rot Pathogen Heteroba sidionannosums.l. and the Biocontrol Fungus Phlebiopsis gigantea. ISBN: 978-91-576-7864-5.

Deshwal, N., Sharma, A.K., Sharma, P. 2011. Review on hepatoprotective plants. Int. J. Pharm. Sci. Rev. Res. 7: 15 – 26.

Dewick, P.M. 2009. Medicinal natural products: a biosynthetic approach. 3rd. ed. Chichester, UK: John Wiley & Sons, Ltd. ISBN 9780470741689.

Ede, S.O., Olaniru, R., Otimenyin, S., Aguiyi, J.C. and Ekwere, E.O. 2012. Analgesic and anti-inflammatory activities of ethanolic extract of mushroom *Ganodermaapplanatum*. *IJRRAS*. 13(1): 349 – 352.

Elkhateeb, W.A., Daba, G. M., Asma, N.E., Sheir, D.H., Fayad, W., Shaheen, M.N., Elmahdy, M. and Wen, T. 2020. Insights into the invitro hypocholesterolemic, antioxidant, antirotavirus, and anticolon cancer activities of the methanolic extracts of a Japanese lichen, *Candelariella vitellina*, and a Japanese mushroom, *Ganoder* maapplanatum. Egyptian Pharmaceutical J. **19:** 67 – 73.

Elkhateeb, W.A., Zaghlor, G.M., Islam, M.E., Ahmed, E.F., Rateb, M.E. and Moneim, A.E.A. 2018. *Ganoderma* applanatum secondary metabolites induced apoptosis through different pathways: In vivo and in vitro anticancer studies. *Biomedicine and Pharmacotherapy*. 101: 064-277.

Elsayed, E.A., Enshasy, H.E., WAdaan, M.A.M. and Aziz, R. 2014. Musrooms: A potential natural source of anti-inflammatory compounds for medical applications. *Mediators of Inflammation*. 805841: 1 – 15.

Ferenbach, D.A. and Bonventre, J.V. 2016. Acute kidney injury and chronic kidney disease: From the laboratory to the clinic. *Nephrol. Ther.* **12(1)**: S41-S48.

First Nature. 2021. Ganoderma applanatum(Pers.) Pat. artist's fungus. https://www.first-nature.com/fungi/ganoderma-applanatum.php

Gao, Y., Tang, W, Gao, H., Chan, E., Lan, J., Li, S., Zhou, S. 2005. Antimicrobial activity of the medicinal mushroom *Ganoderma*. *Food Reviews International*. 21(2): 211 – 229.

Gaoxing, M., Yang, W., Zhao, L., Pei, F., Fang, D. and Hu. Q. 2018. A critical review on the health promoting effects of mushrooms nutraceuticals. Food Science and Human Wellness. 7: 125 – 133. Ginns, J. 2017. *Polypores of British Columbia (Fungi: Basidiomycota).*

Victoria, BC: Province of British Columbia - Forests, Lands, and NR Operations. p. 105. ISBN 978-0-7726-7053-3

Gowda, S., Desai, P.B., Hull, V.V., Math, A.A., Vernerkar, S.N. and Kulkarni, S.S. 2009. A review on laboratory liver function tests. *Pan. Afr. Med. J.* 22: 03 – 17.

Gowda, S., Desai, P.B., Kulkarni, S.S., Hull, V.V., Math, A.A. and Vernekar, S.N. 2010. Markers of renal function tests. *N. Am. J. Med. Sci.* 2(4): 170 173.

Graham, M.L., Janecek, J.L., Kittredge, J.A., Hering, B.J. and Schuurman, H.J. 2011. The Streptozotocin-induced diabetic nude mouse model: differences between animals from different sources. *Comp. Med.* **61(4)**: 356 – 360.

Guzman, M.S., Santafe, G.G., Salcedo, M.M., Angulo, A.A. and Ayaso, O.T. 2013. Chemical study and antioxidant and bactericide activitie from *Ganodermaapplanatum*. *Biotechnologiaenel Sector Agropecuario y Agroindustrial*. 11(1): 88-94.

Hakkim, F.L., Al-Buloshi, M. and Achankunju, J. 2016. Chemical composition and anti-proliferative effect of Oman's *Ganodermaapplanatum* on breast cancer and cervical cancer cells. *J Taibah University Medical Sciences.* **11(2):** 145 – 151.

Halliwel, B. 2007. Biochemistry of oxidative stress. Biochem. Soc.

Trans. 35: 1147-1150.

Hossain, M. S., Barua A., Tanim M. A. H., Hasan M. S., Islam M. J., Hossain M. R., Emon N. U., & Hossen S. M. M. (2021). *Ganoderma applanatum* mushroom provides new insights into the management of diabetes mellitus, hyperlipidemia, and hepatic degeneration: A comprehensive analysis. *Food Science and Nutrition*. **9**: 4364–4374. 10.1002/fsn3.2407

Jeong, Y., Jeong, S., Yang, B., Islam, R. and Song, C. 2009. Optimal culture conditions for mycelial growth and exo-polymer production of *Ganoderma applanatum*. *Mycobiology*. **37(2)**: 89-93.

Kao, C.H.J., Jusuthasan, A.C., Bishop, K.S., Glucinam, M.P. and Ferguson, L.R. 2013. Anti-cancer activities of *Ganodermalucidum*: active ingredients and pathways. Funct. *Foods Health Dis.* **3**: 48 – 65.

Karsten, P. 1881. Enumeration boletinarum et poly porarumfennicarum systemate novo dispositorum. *Revue de Mocologie*. **3(9):** 16 – 18.

Kellum, J.A., Romagnani, P., Ashuntantang, G., Ronco, C., Zarbock, A. and Ander, H.J. 2021. Acute kidney injury. *Nat. Rev. Dis. Primers.* 7(52): 1 – 17.

Klaus, A.S., Kozarski, M.S., Vunduk, J.D., Petrovic, P.M. and Niksic, M.P. 2016. Antibacterial and antifungal potential of wild basidiomycete mushroom *Ganodermaapplanatum*. *Beograd*. **36**: 37 – 46.

Kozarski, M., Klaus, A., Jakovljevic, D., Todorovic, N., Vunduk, J., Petrovic, P., Niksic, M., Virvic, M.V. and Griensven, L.V. 2015. Antioxidants of Edible Mushrooms. *Molecules*. 20: 19489-19525. doi:10.3390/molecules201019489.

Krishnaiah, D., Sarbatly, R. and Nithyanandam R. 2011. A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing.* **8(9):** 217 – 233.

Kumar, D., Arya, V., Kaur, R., Bhat, Z.A., Gupta, V.K. and Kumar, V. 2012. A review of immunomodulators in the Indian traditional health care system. J. Microbiol. Immunol. Infect. 45(3): 165 – 84.

Kuo, M. (2018). Ganoderma applanatum. Retrieved from the *MushroomExpert.Com* Website: http://www.mushroomexpert. com/ganoderma applanatum.html

Lee, C.L., Hung, H.K., Wang, J.J. and Pan. T.M. 2007. Improving the ratio of monacolin K to citrinin production of *Monascuspurpureas* NTU568 under Dioscorea medium through the mediation of pH value and ethanol addition. *J. Agricultural and Food Chemistry*. **55(16)**: 6493-6502.

Li, W.J., Chen, Y., Nie. S.P. et al. 2011. Ganodermaatrum polysaccharide induces anti-tumour activity via the mitochondrial apoptotic pathway related to activation of host immune response. J. Cellular Biochemistry. 112(3): 860 – 871.

Ligouri, I., Russo, G., Curcio, F., Bulli, G., Aran, L., Della-Morte, D., Gargiulo, G., Testa, G., Cacciatore, F., Bonaduce, D. and Abete, P. 2018. Oxidate stress, ageing and diseases. *Clin. Interv. Ageing.* 13: 757-772.

Lin, J.Y. and Chou, T.B. 1984. Isolation and characterization of a lectin from edible mushroom, *volvariellavolvacea*. The J. Biological Chemistry. 96(1): 35 – 40.

Lindequist, U., Kim, H.W., Tiralongo, E. and Griensven, L.V. 2014. Medicinal mushrooms. *Evidence-Based Complimentary and Alternative Medicine*. 1-2: 806180.

Liu, Z., Hou, X., Zhao, J. and He, L. 2015. Liquid fermentation of *Ganodermaapplanatum* and antioxidant activity of exopolysaccharides. *The Open Biomedical Engineering J.* **9**: 190 – 193.

Luo, Q., Wei, X. Y., Yang, J., Luo, J. F., Liang, R., Tu, Z. C. and Cheng, Y. X. 2017. Spiro meroterpenoids from Ganoderma applanatum. J. Natural Products. 80(1): 61–70

Madrigal-Santillan, E., Madrigal-Bujaidar, E., Alvarez-Gonzaliz, I., Sumaya-Martinez, M.T., Gutierrez-Salinas, J., Bautista, M., Morales-Gonzalez, A., Garcia-Luna, Y., Gonzalez-Rubio, M., Aguilar-Faisal, J.L. and Morales-Gonzalez, J.A. 2014. Review of natural products with hepatoprotective effects. *World. J. Gastroenterol.* 20(40): 1487-14804.

Mawar, R., Ram, L., Deepesh, N. A and Mathur, T. 2020 – Ganoderma, In: Amaresan N, Senthil MK, Annapurna K, Kumar K, Sankaranarayanan, A. (eds). Beneficial Microbes in Agro-Ecology. Academic Press. pp. 625–649.

McCarty, M. 2001. Inhibition of acetyl-CoA carboxylase by cystamine may memediate the hypotriglyceridemic activity of pantethine. *Medical Hypothesis*. **56(3):** 314-317.

Meek, I. L. and Vonkeman, H.E. 2010. Nonsteroidal anti-inflammatory drugs: an overview of cardiovascular risks. *Pharmaceuticals*. **3(7)**: 2146-2162.

Mensah, G.A. and Brown, D.W. 2007. An overview of cardiovascular disease burden in the United States. Health Aff. (Mllwood). **26(1):**38-48.

Meuninck, Jim. 2017. Foraging Mushrooms Oregon: Finding, Identifying, and Preparing Edible Wild Mushrooms. Falcon Guides. p. 46. ISBN 978-1-4930-2669-2

Mishra, J., Joshi, A., Rajput, R., Singh, K., Bansal, A. and Misra K. 2018. Phenolic rich fractions from mycelium and fruiting body of *Ganodermalucidum* inhibit bacterial pathogens mediated by generation of reactive oxygen species and protein leakage and modulate hypoxic stress in HEK 293 cell line. *Advances in Pharmacological Sciences*. 6285615: 01 – 10.

Mohammadifar, S., Gharaghoz, S.F., Shayan, M.R.A. and Vaziri, A. 2020. Comparison between antioxidant activity and bioactive compounds of *Ganodermaapplanatum* (Pers.) Pat. And *Ganodermalucidum* (Curt.) P. Karst from Iran. *Iranian J. Plant Physiology*. **11(1)**: 3417 – 3424.

Monika, O., Jaszek, M., Magdalena, M., Blachowicz, A., Rejczak, T. P., Janusz, G., Wydrych, J., Polak, J., Anna, W. and Martyna, K. 2014. Exopolysaccharide from *Ganodermaapplanatum*as a promising bioactive compound with cytostatic and antibacterial properties. *Biomed Research international*. **743812:** 1 – 10.

Moradali, M. 2008. Investigation of antimicrobial fatty acid from medicinal artist conk mushroom *Ganodermaapplanatum* (Pers.) Pat. (aphyllophoromycetidae) by TLC and spectroscopic detection. *International J. Medicinal Mushroom.* **10(2):** 149-154.

Muhsin, T.M., Al-Duboon, A.A. and Khalaf, K.T. 2011. Bioactive compounds from a polypore fungus *Ganodermaapplanatum* (Per s. ex WAllr.) *Pat. Jordan J. Biological Sciences.* **4(4):** 205 – 212.

Nagaoka, S., Shimizu, K., Kaneko, H., Shibayama, F., Morikawa, K., Kanamaru, Y., Otsuka, A., Hirahashi, T. and Kato, T. 2005. A novel protein C-Phycocyanin plays a crucial role in the hypocholesterolemic action of spirulina platensis concentrate in rats. *The J. Nutrition*. **135(10)**:2425-2430.

Nagaraj, K., Mallikarjun, N., Naika, R. and Venugopal, T.M. 2013. Phytochemical analysis and *in-vitro* antimicrobial potential of *Ganodermaapplanatum* (Pers.) Pat. Of Shivamogga district-Karnataka, India. Int. J. Pharm. Sci. Rev. Res. 23(2): 36 – 41.

Nagaraj, K., Mallikarjun, N., Naika, R., Venugopal, T.M. 2014. Antioxidative activities of wild macro fungi *Ganodermaapplanatum* (Pers.) *Pat. Asian J. Pharmaceutical and Clinical Research*. **7(2):** 166– 171.

Packard, C.J., Boren, J. and Taskinen, M.R. 2020. Causes and consequences of hypertriglyceridemia. *Frontiers in Endocrinology*. 11: 252.

Palanna, K.B., Shreenivasa, K.R., Basavaraj, S. and Narendrappa, T. 2020. Review of genus *Ganoderma* causing basal stem rot (coconut) and food rot (arecanut) with respect etiology and management. International *J. Current Microbiology and Applied Sciences.* 9(4): 1434 – 1455.

Parkin, D. M. 2006. The Global health burder of infection-associated cancers in the year 2002. *Int. J. Cancer.* **118:** 3030e3044.

Patel, S. and Goyal, A. 2012. Recent developments in mushrooms as anti-cancer therapeutics: a review. *Biotech.* 2: 1-15.

Paterson, R. R. M. 2006. Ganoderma – A therapeutic fungal biofactory. *Phytochemistry*. 67: 1985 – 2001.

Paul M. Kirk., Paul F. Cannon., David W. Minter and J. A. Stalpers. 2008. Dictionary of the Fungi. 10th Edition. CABI, UK, J Stalpers, CBS, The Netherlands. ISBN: 9780851998268

Pizzino, G., Irrera, N., Cucinotta, M., Pallio, G., Mannino, F., Arcoraci, V., Squadrito, F., Altavilla, D. and Bitto, A. 2017. Oxidative stress: Harms and benefits for human health. Oxid. Med. Cell. Longev. 8416763.

Pranav Kumar and Usha Mina, 2019. Life sciences: Fundamental and practices. Pathfinder publication, New Delhi, ISBN: 978-81-906427-7-4. PP.319-320.

Qureshi, S., Pandey, A. K. and Sandhu, S.S. 2010. Evaluation of antibacterial activity of different *Ganodermalucidum* extracts. *People's J. Scientific Research.* **3(1):** 9 – 13.

Rahman, S. and Parvin, R. 2014. Therapeutic potential of Aegle marmelos (L.) – An overview. Asian Pac. J. Trop. Dis. 4(1): 71-77.

Rajoriya, A., Tripathy, S.S. and Gupta, N. 2015. *In-vitro* antioxidant activity of selected *Ganoderma* species in Odisha, India. *Tropical Plant Research*. 2(2): 72 – 77.

Rasheed, A., Fathima, R. and Azeez, A. 2019. A review on natural antioxidants. Edited by: Cengiz Mordeniz, Traditional and Complementary Medicine. DOI: 10.5772/intechopen.82636

Rathore, H., Prasad, S., Sharma, S. et al. 2017. Mushroom nutraceuticals for improved nutrition and better human health: A review. *Pharma. Nutr.* 5:35 – 46.

Rees, T.M. and Brimijoin, S. 2003. The role of acetylcholinesterase in the pathogenesis of Alzheimer's disease. *Drugs Today (Barc)*. **39(1):**75 – 83.

Rikka, L., Dhanik, R., Pyri, V., Marta, C., Henri, V., Varpu, M. 2018. Antiviral agents from fungi: diversity, mechanisms and potential applications. *Frontiers in Microbiology*. 9: 23-25.

Ruby, V., Dalvi, Y.B., Nai, C.K.K. and Vaidyanathan, K. 2018. Toxicological evaluation and oral glucose tolerance test of *Ganodermaapplanatum* (Pers.) Pat. From Kerala. RJLBPCS. **4(4):** 391 – 412.

Saha, R., Dutta, A.K., Paloi, S., Roy, A. and Acharya, K. 2018. Contribution to the Macromycetes of West Bengal, India. J. Threatened Taxa. 10(15): 13006-13013.

Saify, Z. S. and Sultana, N. 2014. Role of acetylcholinesterase Inhibitors and Alzheimer Disease. In: Atta-ur-Rahman, Muhammad Iqbal Choudhary, Eds., Drug Design and Discovery in Alzheimer's Disease. ISBN 9780128039595, Elsevier. Pp. 387-425. https://doi.org/ 10.1016/B978-0-12-803959-5.50007-6.

Schaiff, R.A., Moe, R.M. and Krichbaum, D.W. 2008. An overview of cholesterol management. Am. Health. Drug. Benefits. 1(9):39-48.

Schmeltz, L. and Metzger, B. 2006. Diabetes/Syndrome X. In: John B. Taylor, David J. Triggle EDs. Comprehensive Medicinal Chemistry II, Elsevier, ISBN 9780080450445, pp.417-458, https://doi.org/10.1016/B0-08-045044-X/00179-6

Seto, M., Honma, K. and Nakagawa, M. 2010. Diversity of genome profiles in malignant lymphoma. *Cancer Sci.* 101: 573e578.

Shaikh, P. Z. 2011. Cytokines & their physiologic and pharmacologic functions in inflammation: *A review. Int. J. Pharm. Life. Sci.* 2: 1247 – 1263.

Shattat, G. F. 2014. A review article on hyperlipidemia: types, treatments and new drugs targets. *Biomedical and Pharmacology J.* **7(2):** 399-409.

SUKUMAR DANDAPAT et al.,

Siegel, R.L., Miller, K.D. and Jemal, A. 2016. Cancer statistics. CA *Cancer J. Clin.* 66: 7 - 30.

Singh, N., Kesherwani, R., Tiwari, A.K and Patel., D.K. 2016. A review on diabetes mellitus. *The Pharma Innovation J.* 5(7): 36 – 40.

Singh, R., Singh, A.P., Dhingra, G.S. and Shri, R. 2014. Taxonomy, physicochemical evaluation and chemical investigation of *Ganoderma* applanatum and *G. brownii. International J. Advanced Research.* **2(5):** 702 – 711.

Sinha, M., Gautam, L., Shukla, P.K., Kaur, P., Sharma, S. and Singh, P.T. 2013. Current perspectives in NSAID-induced gastropathy. Mediators of Inflammation. 258209: 1-11.

Smania, A., Monache, F.D., Smania E.F.A. and Cuneo, R.S. 1999. Antibacterial activity of steroidal compounds isolated from *Ganodermaapplanatum* (Pers.) Pat. (Aphyllophoromycetideae) fruit body. *International J. Medicinal Mushroom*. 1(4): 325 – 330.

Stern, M.P., Williams, K. and Haffner, S. M. 2002. Identification of persons at high risk for type 2 diabetes mellitus: do we need the oral glucose tolerance test? *Annals of Internal Medicine*. **136**: 575 – 581.

Stumvoll, M., Mitrakou, A., Pimenta, W., Jenssen, T., Yki-Jarvenin, H.A. and Van. H.T. 2000. Use of the oral glucose tolerance test to assess insulin release and insulin sensitivity. *Diabetes Case*. 23: 295-301.

Valko M., Leibfritz, D., Monocola, J., Cronin, M.D., Mazur, M., Telser, J. 2007. Free radicals and antioxidants in normal physiological functions and human disease. *The International J. Biochemistry & Cell Biology.* **39**: 44 – 84.

Valko, M., Rhodes, C.J., Monocol, J., Izakovic, M., Mazur, M. 2006. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*. **160**: 1 – 40.

Wachtel-Galor, S., Yuen, J., Buswell, J. A. et al. 2011 Ganoderma lucidum (Lingzhi or Reishi): A Medicinal Mushroom. In: Herbal Medicine: Biomolecular and Clinical Aspects. Benzie, I. F. F. and Wachtel-Galor, S., (Eds). 2nd edition. Boca Raton (FL): CRC Press/ Taylor & Francis; 2011. Chapter 9. Available from: https:// www.ncbi.nlm.nih.gov/books/NBK92757/

Wang, F., Dong, Z. and Liu, J. 2007. Benzopyran-4-one derivatives from the fungus *Ganodermaapplanatum*. Zeitschrift fur Naturforschung B. **62(10)**: 1329-1332.

Wasser, S. P, Coates P, Blackman M, Cragg G, Levine M, Moss J, White J. Encyclopedia of Dietary Supplements. New York: Marcel Dekker; 2005. Reishi or Lingzhi (*Ganoderma lucidum*) pp. 680–90. Woo-Sik, J., Yun-Ju, C., Doo-Hyun, C., So-Deuk, P., Young-Bok, Y. and Seok, S. 2009. Culture conditions for mycelian growth of *Ganoderma applanatum*.*Mycobiology*. **37(2):** 94 – 102.

Xinling, S., Weijun, C., Zheng, G., Jianjun, Z. and Jia, L. 2021. Structural characterization and amelioration of sulphated polysaccharides from *Ganodermaapplanatum* residue against CCl₄induced hepatotoxicity. *International Immunopharmacology*. **96**: 107554.

Yan, L.J. 2014. Positive oxidative stress in ageing and ageing-related disease tolerance. *Redox Biology*. 2C: 165 – 169.

Yang, B.K., Jung, Y.S. and Song, C.H. 2007. Hypoglycemic effects of *Ganodermaapplanatum* and *Collybiaconfluens* exopolymers in streptozotocin-induced diabetic rats. *Phytother. Res.* 21: 1066-1069.

Yang, B.K., Kim, D.H., Jeong, S.C., Das, S., Choi, Y.S., Shin, J.S., Song, S.C. and Song. C.H. 2002. Hypoglycemic effect of *Lentinus* edodesexo-polymer produced from a submerged mycelial culture. *Bioscience Biotechnology and Biochemistry*. **66(5):** 937 – 942.

Yong, T., Chen, S., Xie, Y., Chen, D., Su, J., Shuai, O., Jiao, C. and Zuo, D. 2018. Hypouricemic effects of *Ganodermaapplanatum* in hyperuricemia mice through OAT1 and GLUT9. *Front. Pharmacol.* 8:996.

Yong-Tae, J., Byung-Keun, Y., Sang-Chul, J., Sang-Min, K. and Chi-Hyun S. 2008. *Ganodermaapplanatum*: a promising mushroom for zntitumor and immunomodulating activity. Phytotherapy research. 22(5): 614-619.

Yu. C., Xiaowei, x., Shujing, J., Linfang, H. and Jian, G. 2018. *Ganoderma*: A cancer immunotherapy review. *Front. Pharmacol.* 9:1217.

Yuandani, Ibrahim, J., Sri, R.A. and Bagus, S.I. 2021. Immun omodulatory effects and mechanisms of curcuma species and their bioactive compounds: a review. *Frontiers in Pharmacology*. **12**: 643119.

Zengin, G., Sarikurkcu, C., Gunes, E., Uysal, A., Ceylan, R., Uisal, S., Gungor, H. and Aktumsek, A. 2015. Two Ganoderma species: profiling of phenolic compounds by HPLC-DAD, antioxidant, antimicrobial and inhibitory activities on key enzymes linked to diabetes mellitus, Alzheimer's disease and skin disorders. *Food Funct.* **6**:2794-2802.

Zhang, L., Xu, S., Liang, W., et al. 2015. Antibacterial activity and mode of action. *Tropical J. Pharmaceutical Research*. 14(11): 2099-2106.